

# Forced Degradation of Proteins Through Mechanical Stress



**Fluence** Analytics

realtime data. realtime optimization.

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# ARGEN

- Continuously measures light scattering from 16 independently Temperature and Stirring controlled cells
- Early detection of aggregation and degradation
- Key Outputs
  - Aggregation Rate (AR)
  - Normalized Mw
  - Mass AR
- Applications
  - Formulation stability
  - Interfacial studies
  - Material interaction
  - GPC scheduling
  - Dissolution kinetics

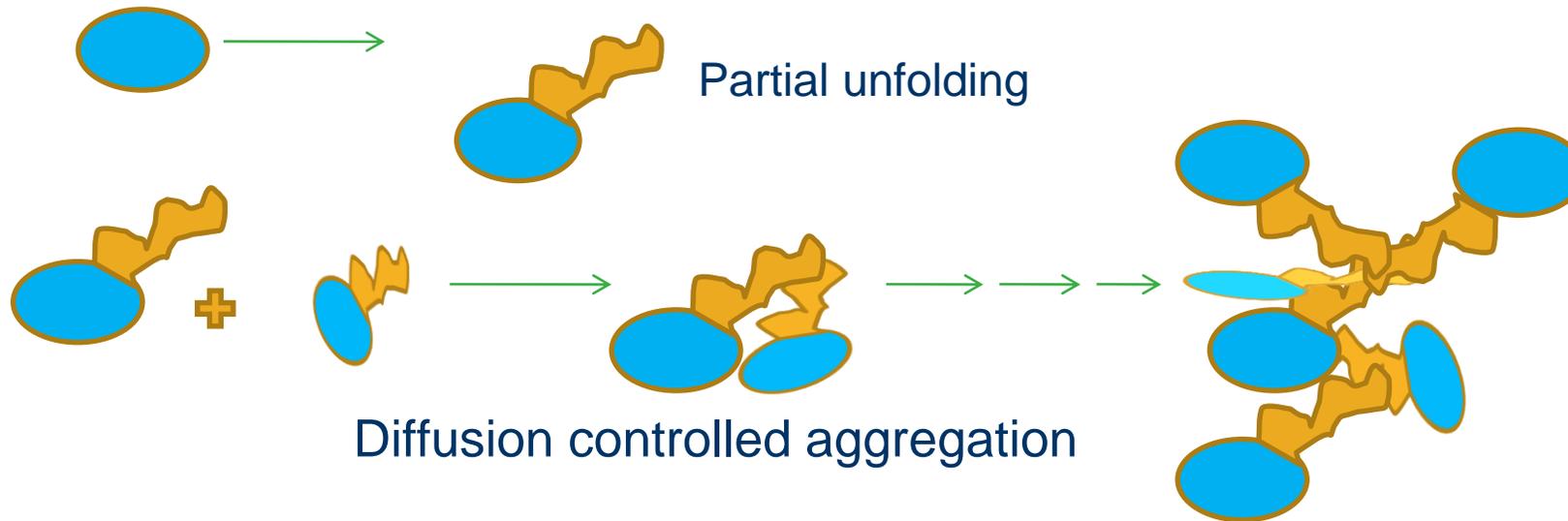


## Specifications:

- Temp Control: 18 – 100 C
- Stirring Control: 0-2000 RPM
- Power: 90 – 220 VAC
- Dimensions: 30" w x 22" d x 14" h
- Weight: 85 lbs

# Protein Aggregation

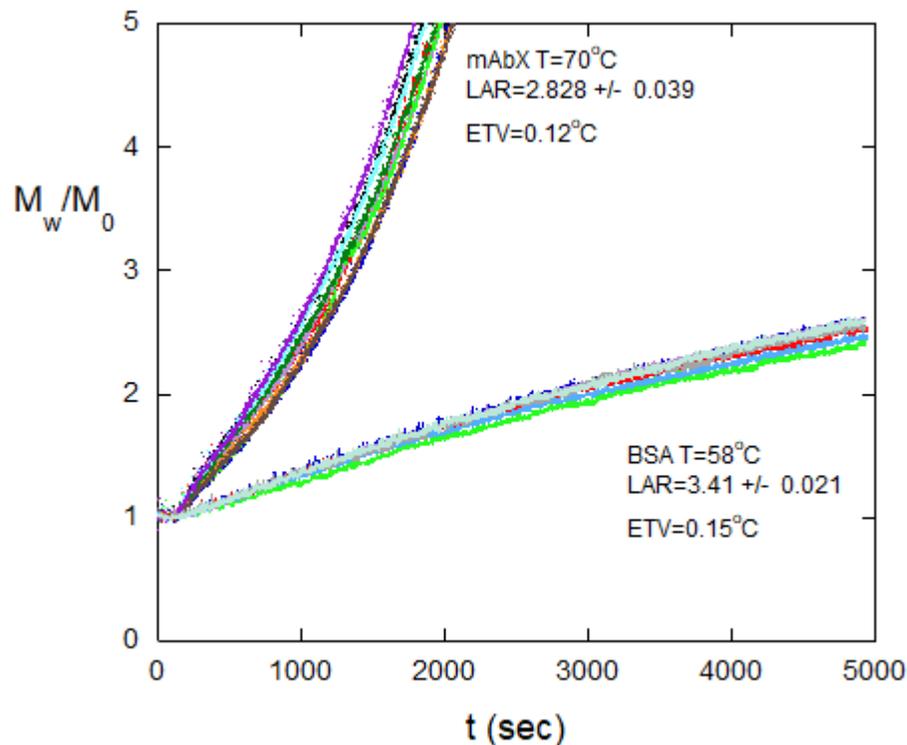
Native Protein



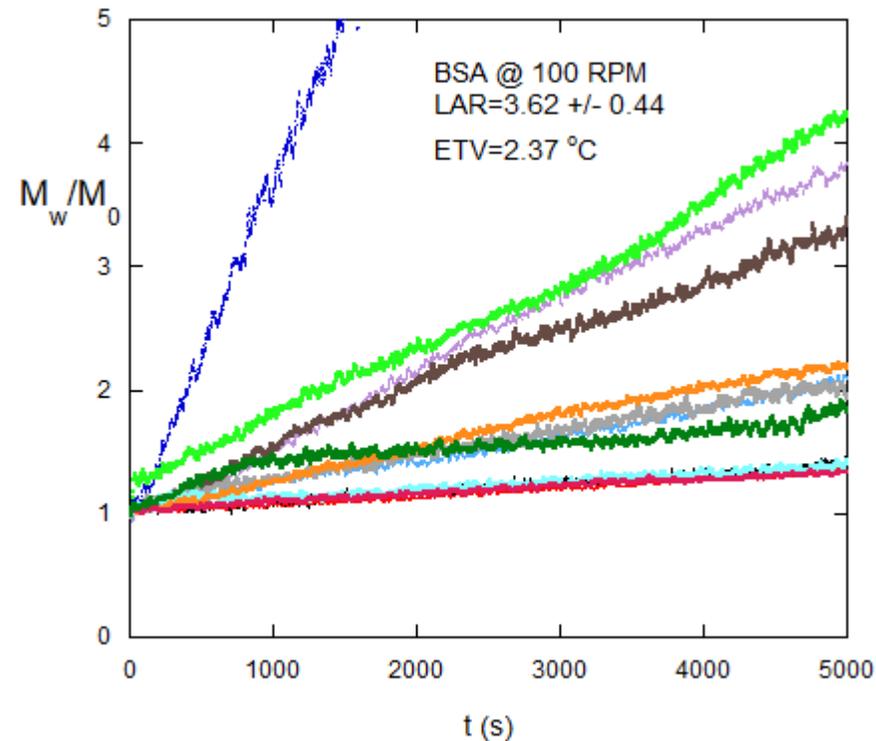
- Major problem in development of new therapeutic proteins
- Several mechanisms exist, highly protein dependent
- Need for high throughput screening

# Routine Aggregation Monitoring via ARGEN

## Routine Methods:



Thermally induced aggregation



Contact-stir induced aggregation

# Non-Routine Aggregation Monitoring via ARGEN

## Mechanical/Shear Stress

- Non-contact stirring with overhead suspended magnetic stir bars controlled by the ARGEN software (3d-printed construction with inexpensive and readily available components)
- Examination of capillary shear through different tubing diameter and composition via reciprocating syringe pump
- Non-contact stirring by overhead impellers controlled by stepper motors (motors and drivers are inexpensive and readily available)
- Recirculation of protein samples from sample cuvette via peristaltic pump
- Recirculation of protein samples through differing filter materials via peristaltic pump
- Continuous extraction of protein samples from ARGEN through a secondary detector (MALS, fluorescence, UV, etc.) via syringe to provide complimentary analytical methods
- Continuous or discrete extraction of protein samples for GPC analysis with an autoinjector

## Non-Mechanical/Shear

- Direct titration in real time with salts, excipients, acids/bases, denaturation agents, etc.
- Generation of Debye plots
- Probing Host:Guest interactions

# Example of Equipment for Non-Routine Experiments

## Mechanical/Shear Stress

- 3-d printer and/or basic machining equipment (overhead stirring)
- A good source for small parts for stirring is McMaster-Carr, <https://mcmaster.com>
- Syringe pump (capillary shear stress, withdrawal through secondary detectors)
- Peristaltic pump (recirculation, filtration)
- HPLC tubing and various fittings

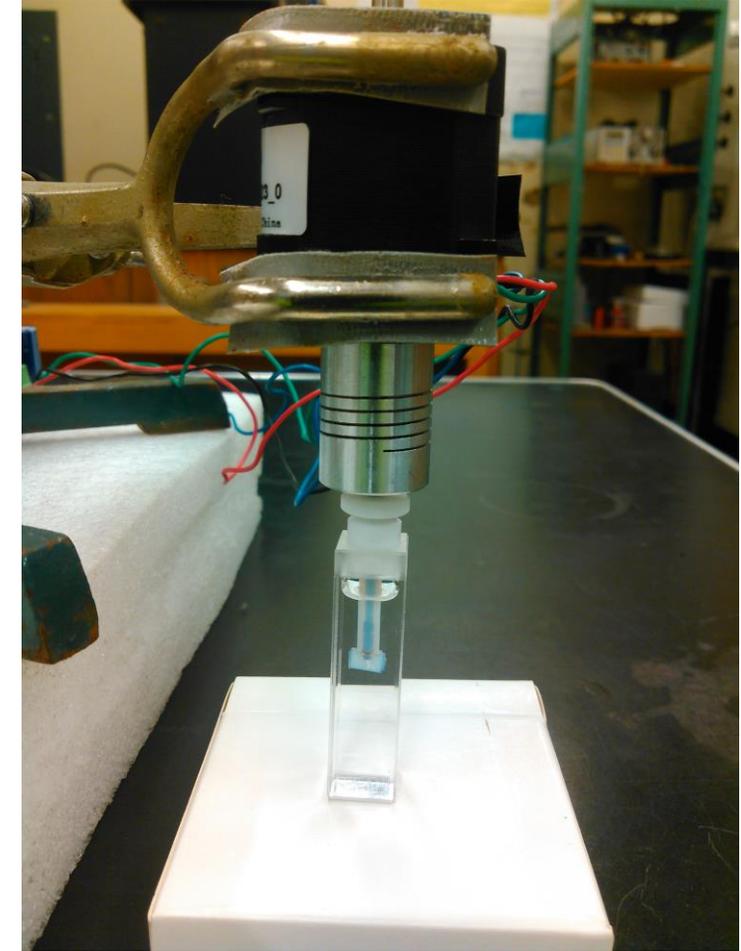
## Non-Mechanical/Shear

- Non-routine applications such as titrations and Debye plots are easier to analyze using the data export function, generates .csv file for external analysis

# Non-Contact Stirring



- Can be coupled to instrument control (left) or independently controlled via stepper motor (right)
- Stirrer consists of magnetic stir bar connect to stainless steel or PTFE shaft, miniature bearings pressed into cap (instrument control) or directly inserted through a drilled PTFE cap (stepper motor controlled)
- Shaft/stirrer must not block optical path (~3-4 mm from bottom of cell)

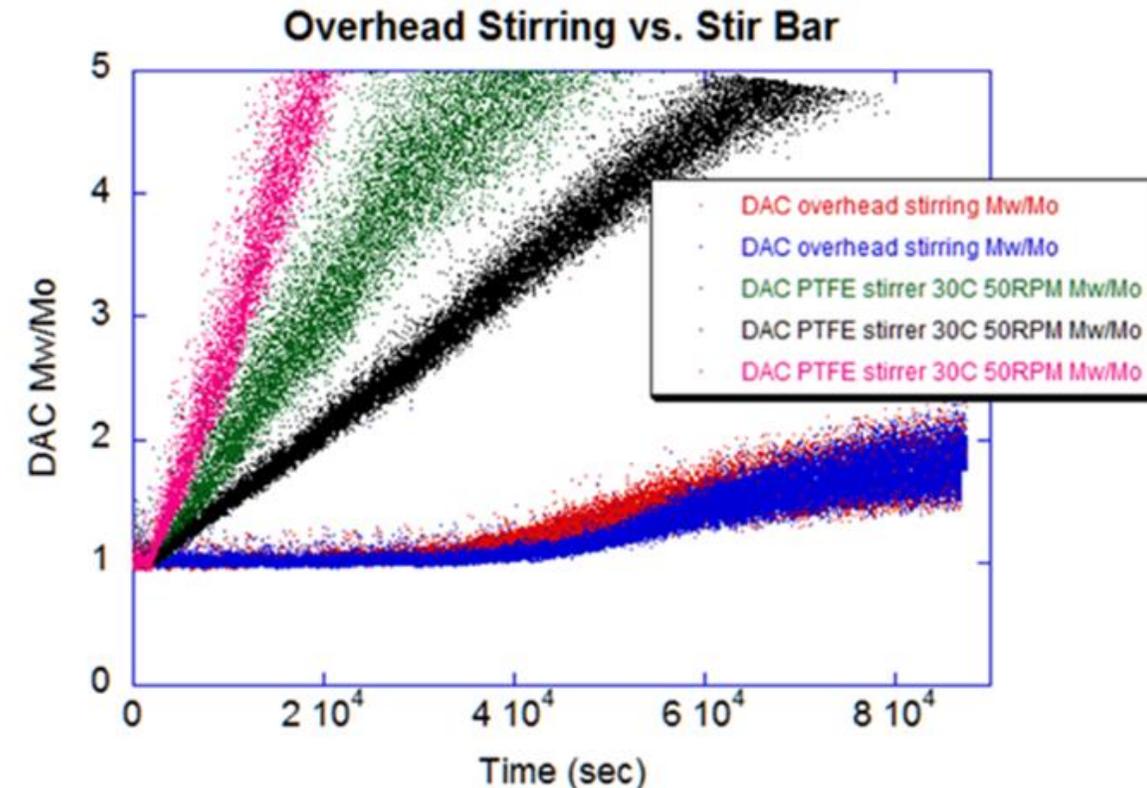


# Overhead vs. Contact Stirring

- Overhead stirring results in markedly decreased aggregation
- Proteins examined to date have demonstrated vastly improved reproducibility relative to contact stir
- Lag phases are commonly observed

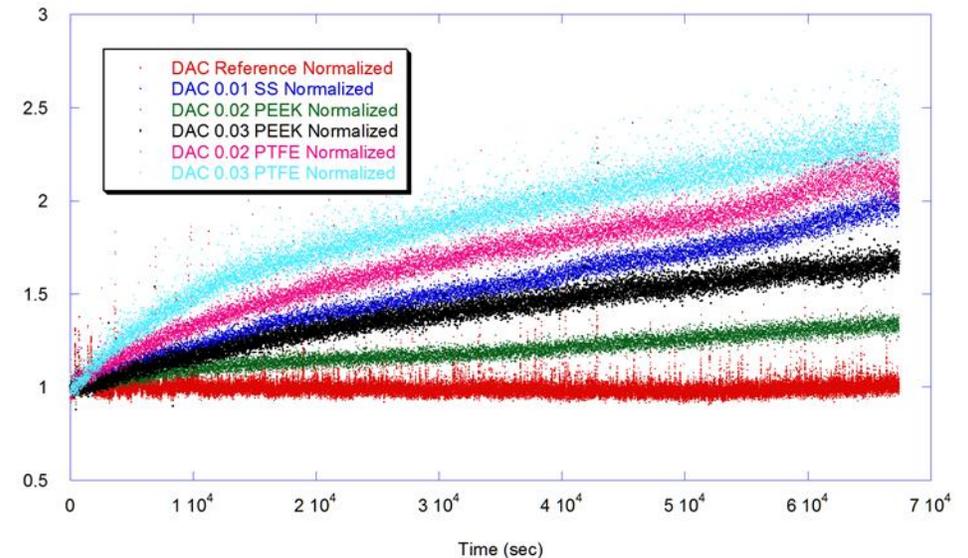
Note:

- Coupled magnetic stirring suitable for samples which aggregate at <500 rpm, stepper motor control required for higher rpm



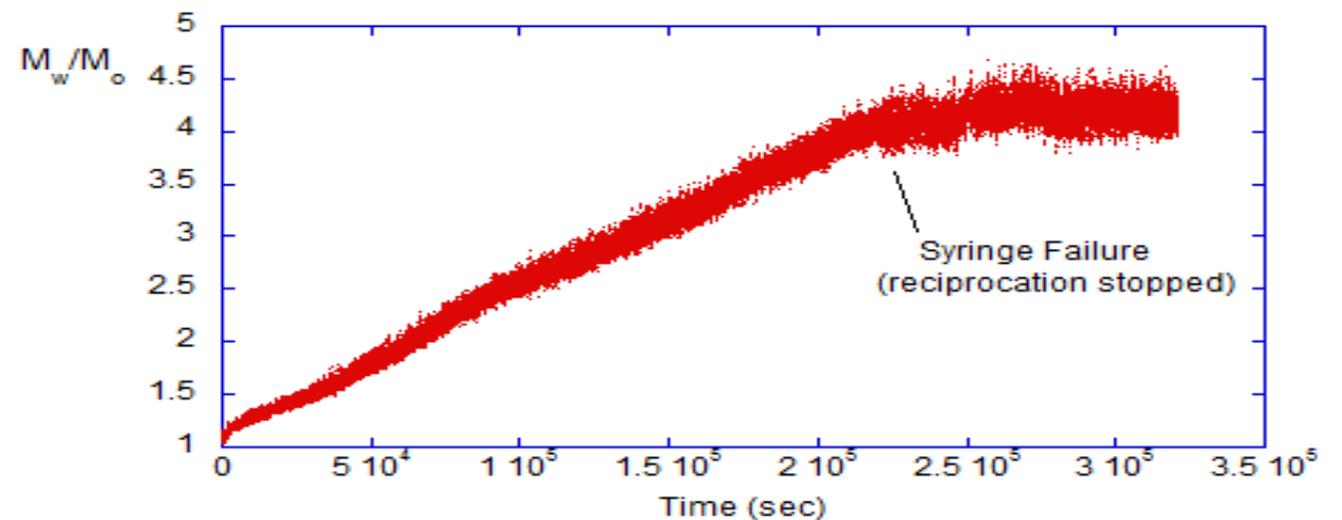
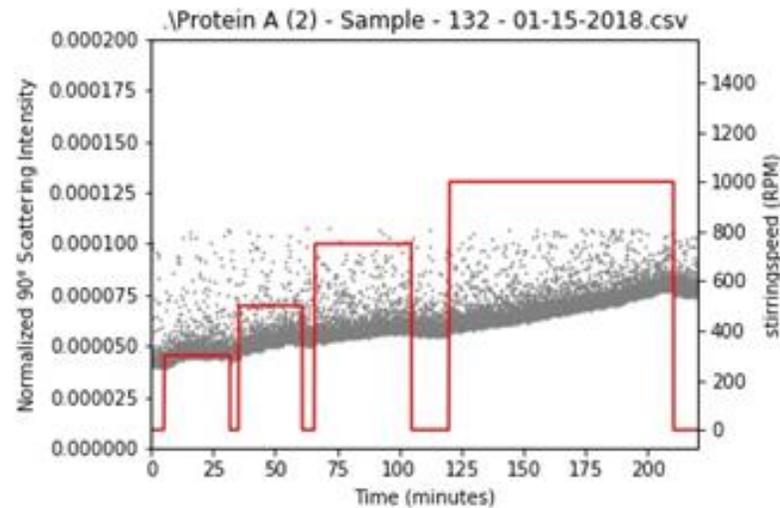
# Capillary Shear Stress

- Utilizes a reciprocating syringe pump to continually extract/inject samples into cuvette under ARGENT monitoring
- Equal lengths of tubing used (1m is sufficient)
- Equal starting volumes of samples required for direct comparison
- Consistent flow rates used
- Tubing material and diameter can have a significant impact on observed aggregation rates, for samples shown in the plot the AR as a function of tubing material is PTFE>SS>PEEK



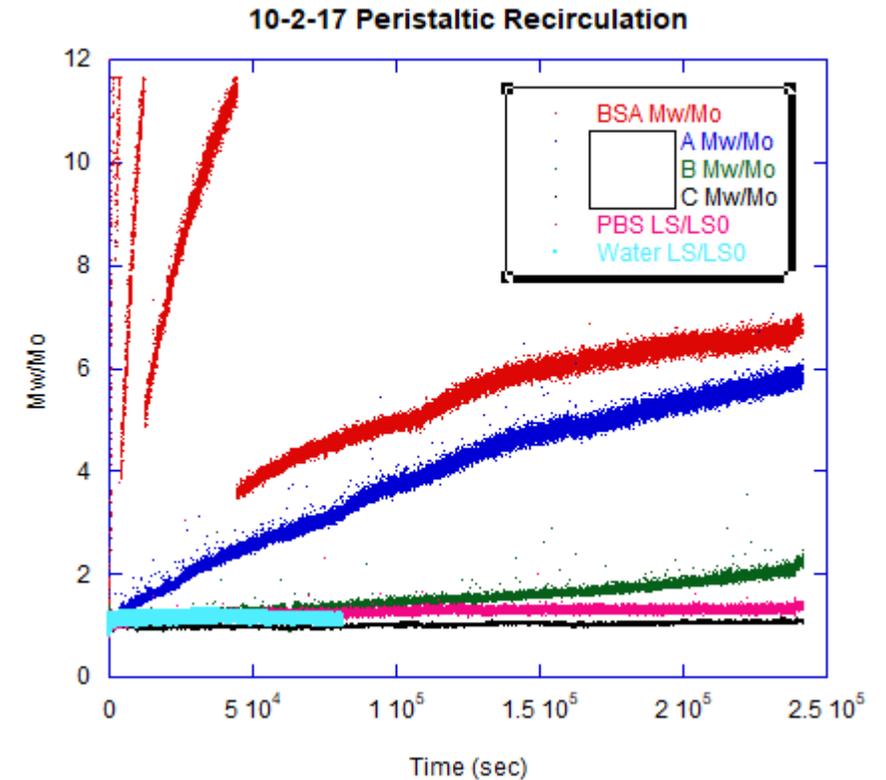
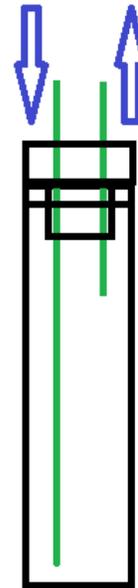
# Stir and Shear Stress Aggregation Halt when Stress Removed

- Aggregation stops when stressor removed
- Upon re-application of stress, aggregation will continue along a similar path



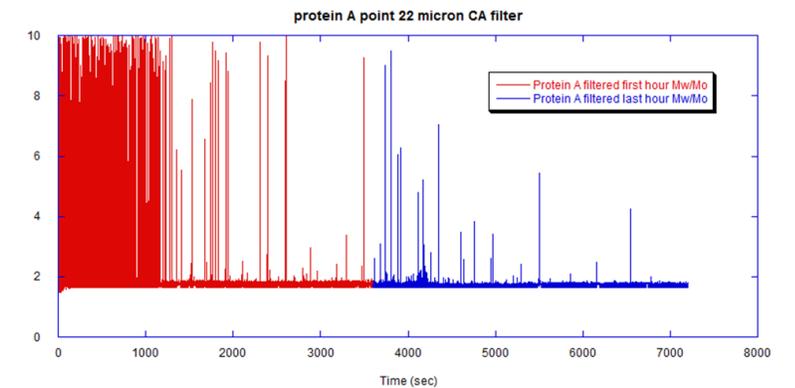
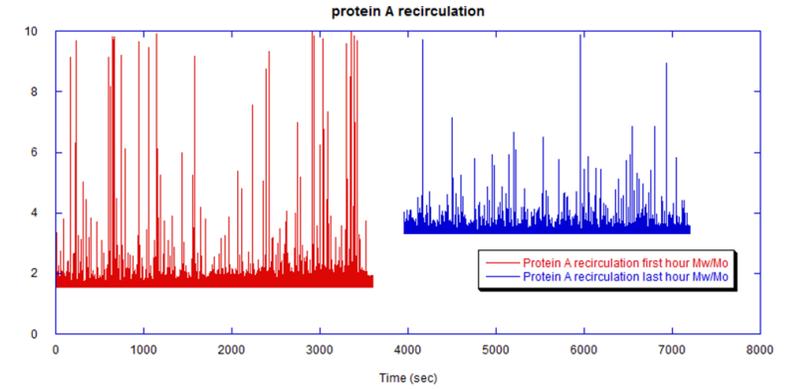
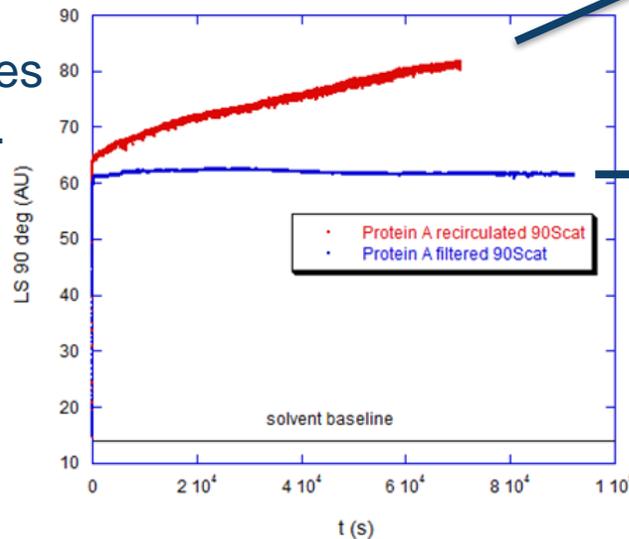
# Peristaltic Recirculation

- Peristaltic pumping generally considered to exert minimal shear stress
- Sample recirculated through cuvette by HPLC tubing
- Water and buffer controls show no increase in light scattering
- The stability of differing proteins varies widely



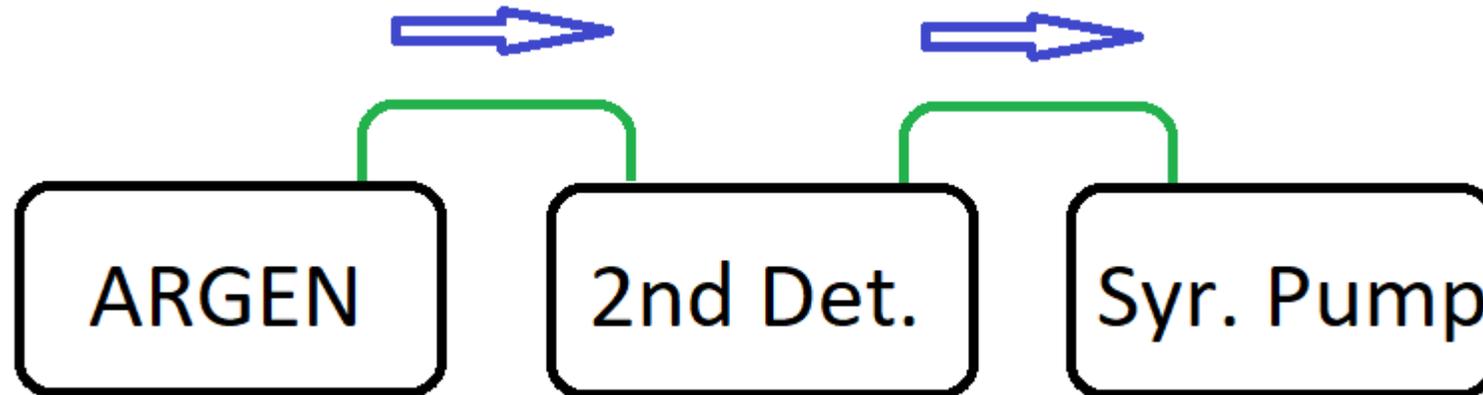
# Peristaltic Recirculation w/ Filtration

- Syringe filter placed in recirculation loop via luer lock fittings
- Filtered sample shows no increase in light scattering and a decrease in scattering spikes caused by large aggregates
- Behavior indicative of a small population of large aggregates leading to increased light scattering signal- SLS intensity is a function of the average population, i.e. a small population of large aggregates can yield an increase similar to a large population of small aggregates
- Confirmation of mechanism requires secondary detection methods, e.g. GPC or MALS

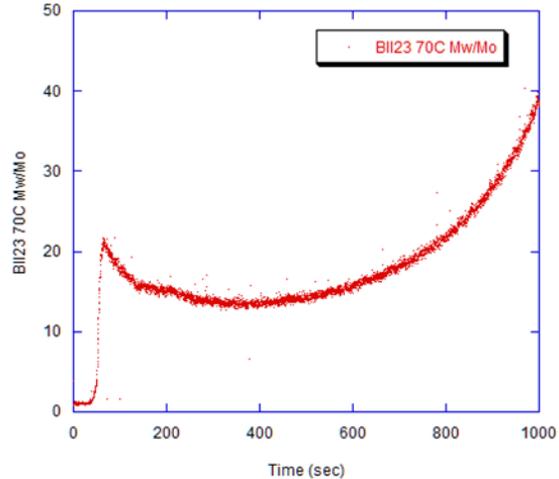


# Coupling to Secondary Detector

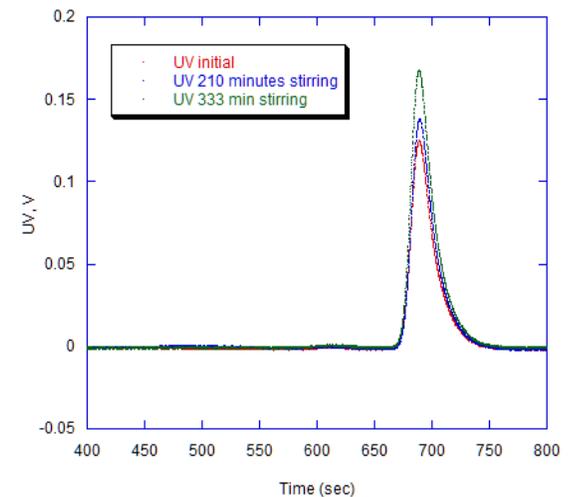
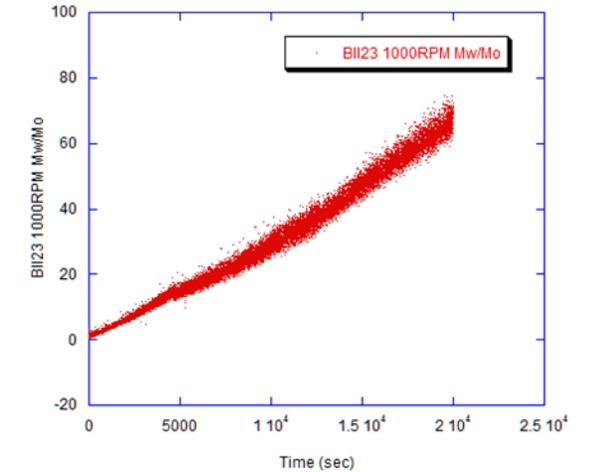
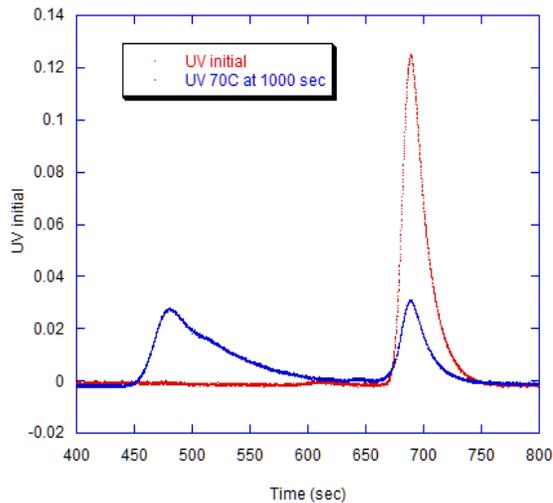
- Syringe pump used to withdraw sample via HPLC tubing
- Can be withdrawn directly through detector (MALS, UV, Fluorescence, etc.)
- Can be coupled to autoinjector for GPC (Rheodyne MXP7900)
- Begin pulling sample before stressor applied
- Minimize dead volume from ARGEN to secondary detector



# Analysis of Thermal vs Stir Stress by Coupled GPC

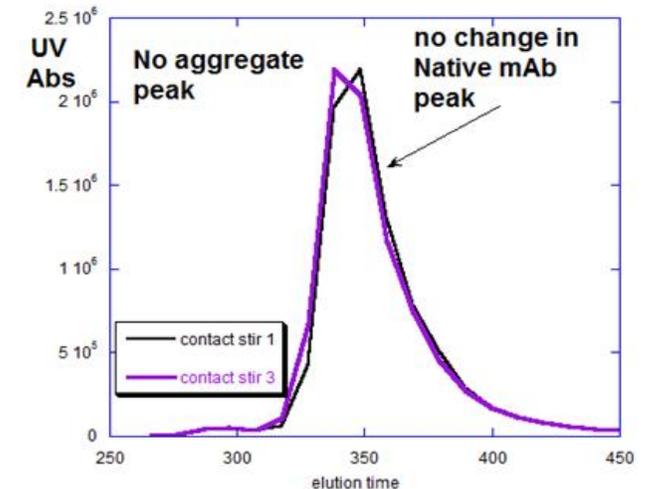
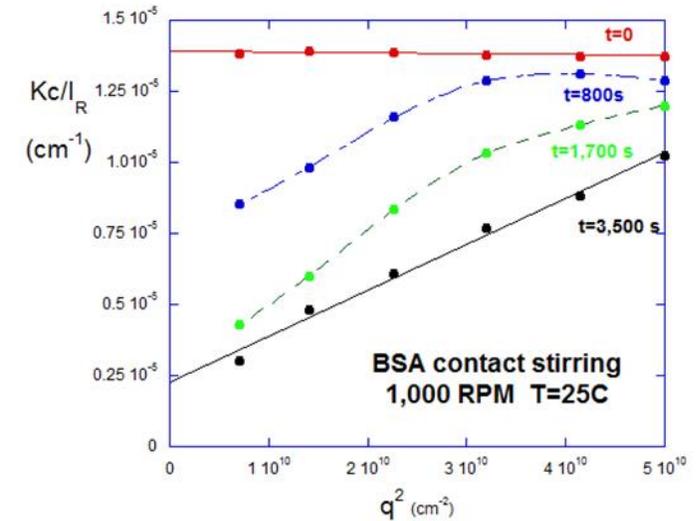
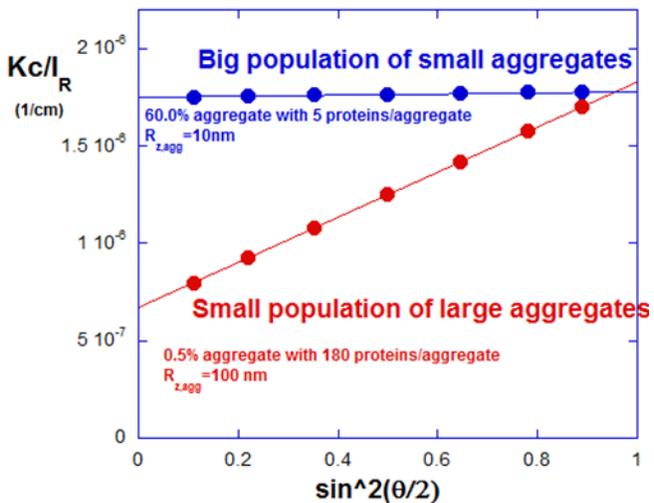


- Sample withdrawn from cuvette through an autoinjector via syringe pump
- Aliquots periodically injected onto GPC column w/UV detection
- Significant monomer loss and aggregate formation observed in thermally stressed sample (left) but not in contact-stir stressed sample (right)

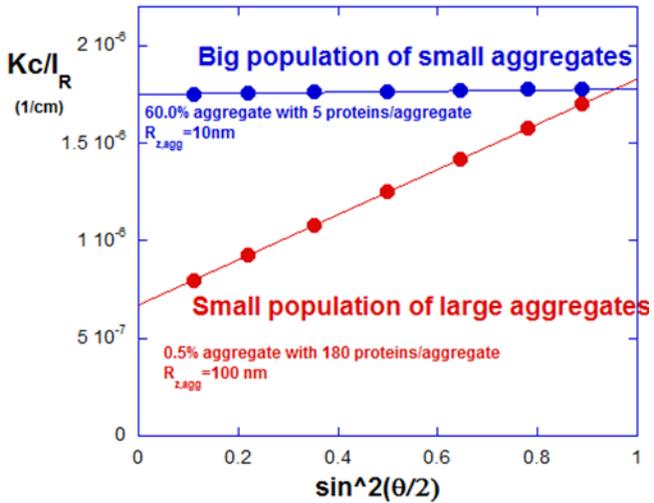


# Analysis of Stir-Stress by Coupled ARGENT-GPC-MALS

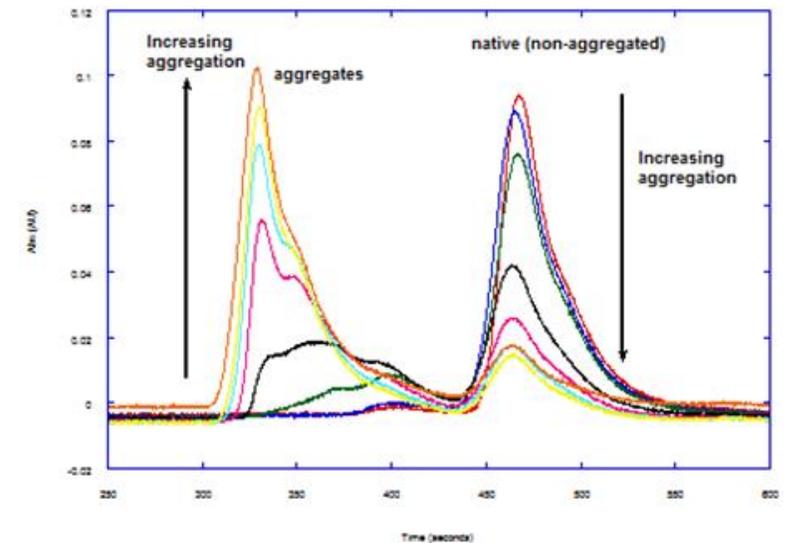
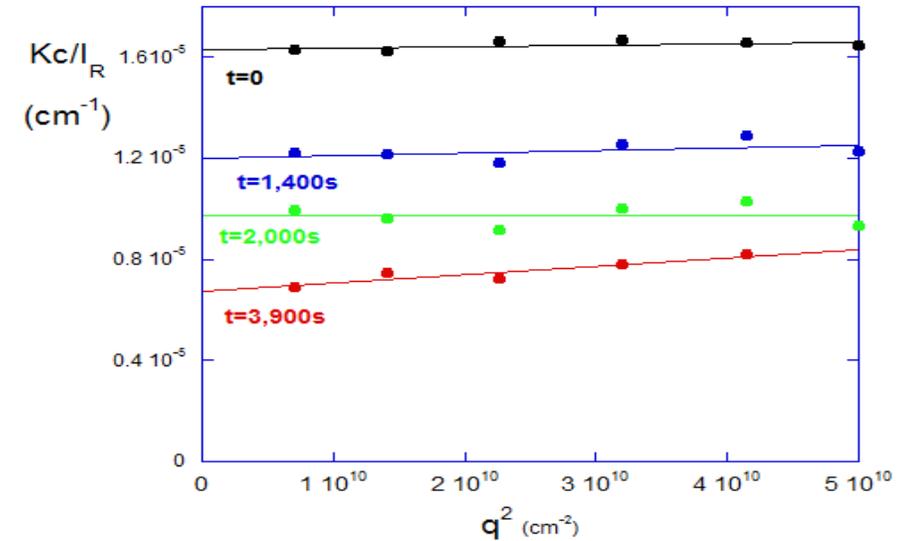
- MALS expands ARGENT capability to discern the nature of aggregate populations
- Angular dependence increases with aggregate size
- Can discern between a big population of small aggregates (thermal stress) and a small population of large aggregates (stir stress)



# Analysis of Thermal Stress by Coupled ARGON-GPC-MALS

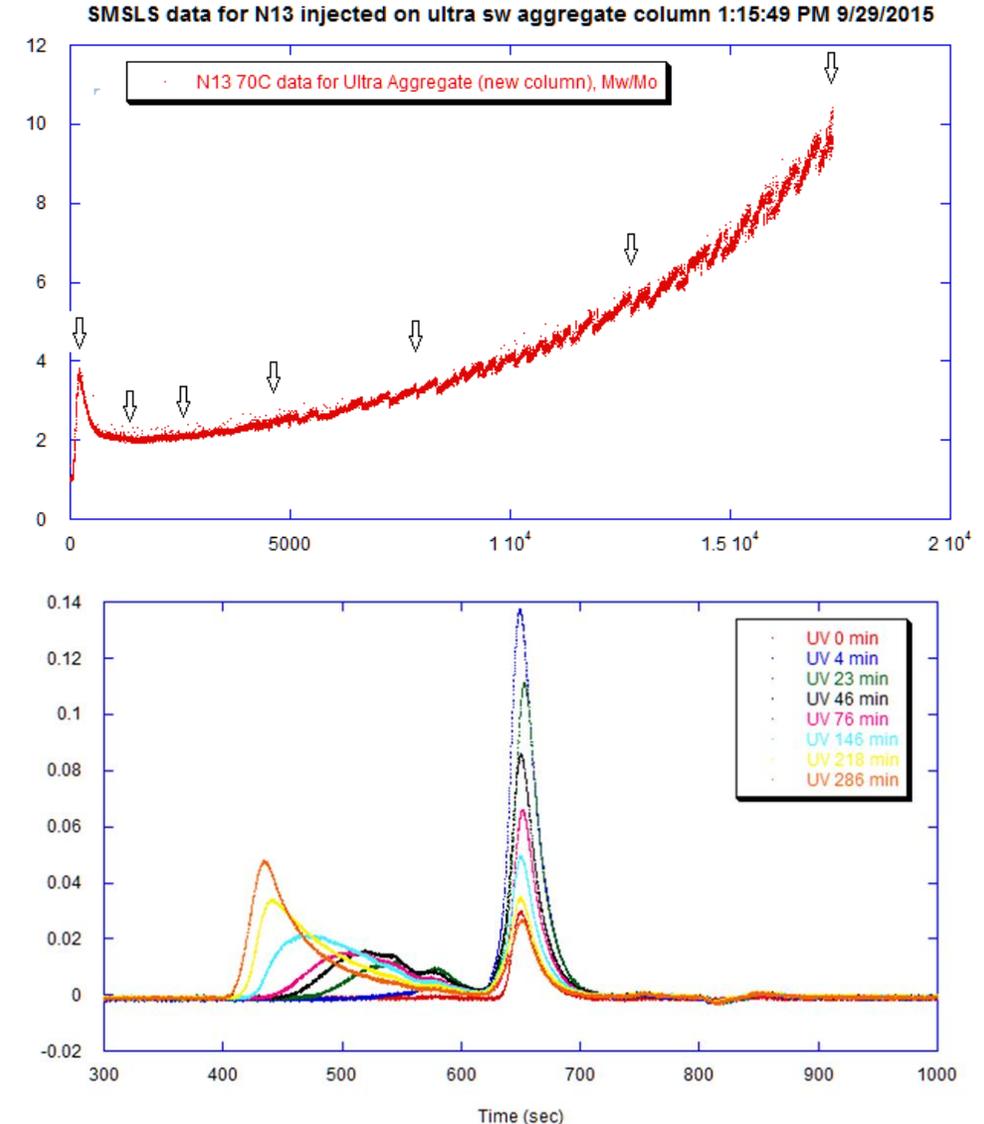


- In contrast to stir stress, thermal stress yields a very small change in angular dependence
- MALS data in agreement with GPC data showing small aggregate formation during thermal stress



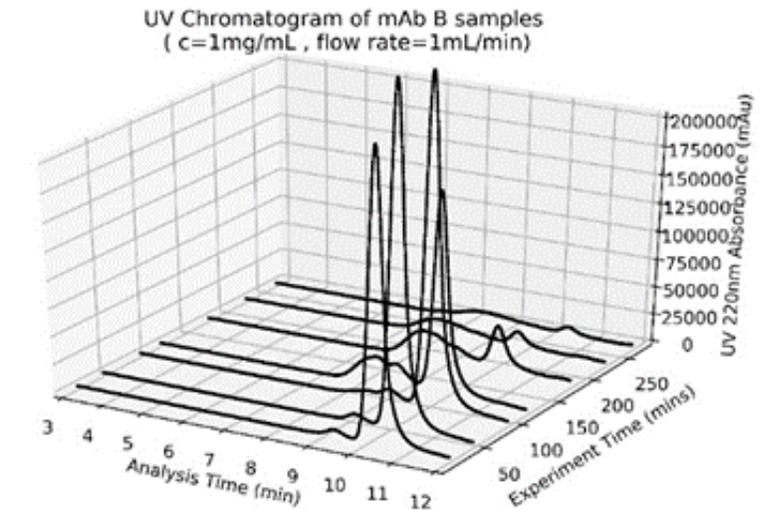
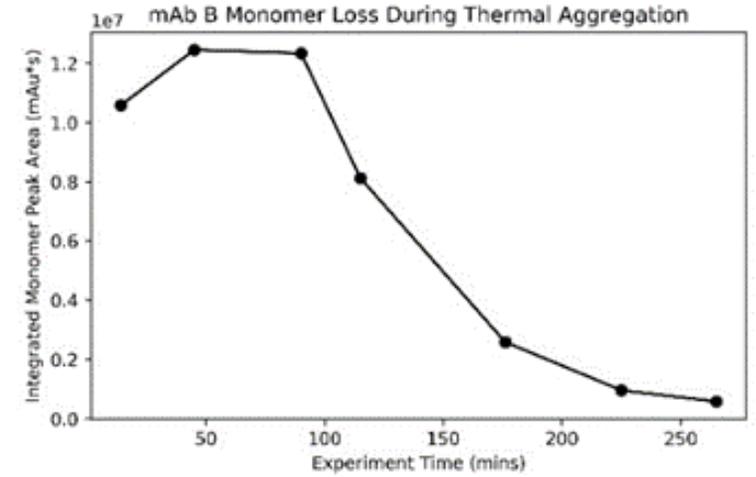
# Observing Aggregate Formation by Coupled GPC

- Thermally stressed sample monitored simultaneously by ARGEN and coupled GPC with a Tosoh Ultra SW aggregate column
- Stepwise aggregation is observed which in which the sample can appear to undergo an evolution through dimeric, trimeric, etc aggregate formation
- Mechanically (contact-stir) stressed samples do not exhibit this same evolution



# Protein Monomer Loss by Coupled GPC

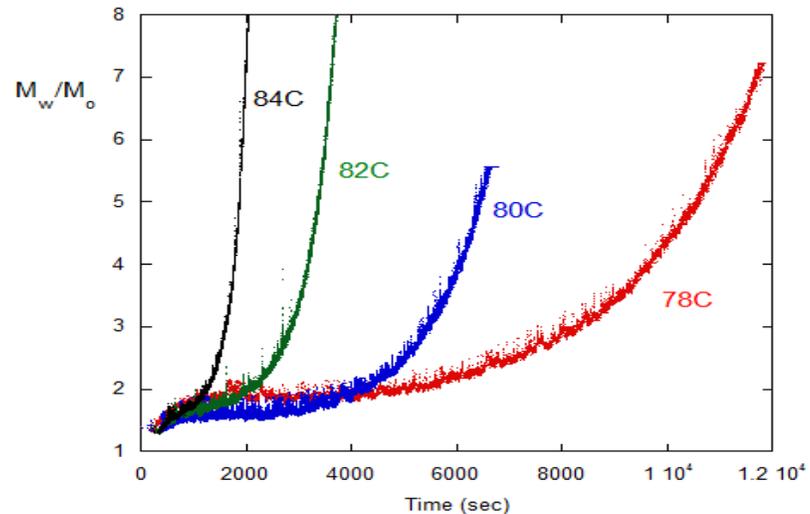
- Protein monomer loss can be calculated with a standard GPC column if an aggregate-specific column not available
- Monomer loss quantified relative to initial area of peak prior to application of stress by ARGENT



# Superposability of LS Signatures Implies the Same Mechanism

**Hypothesis:** If there exists a proportionality constant between the time bases of non-linear  $I_s(t)$  from two aggregation processes which yields an exact superposition of the non-linear  $I_s(t)$  then the kinetic pathways for the two processes are of the same kind.

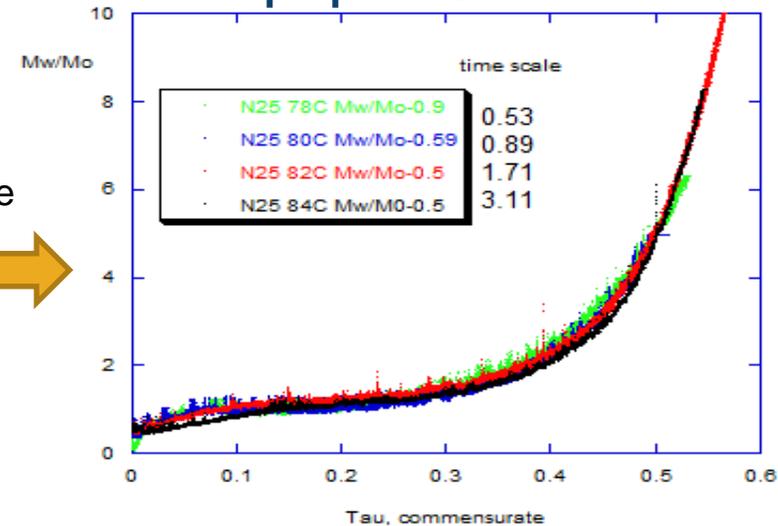
Thermal stress: Real time data



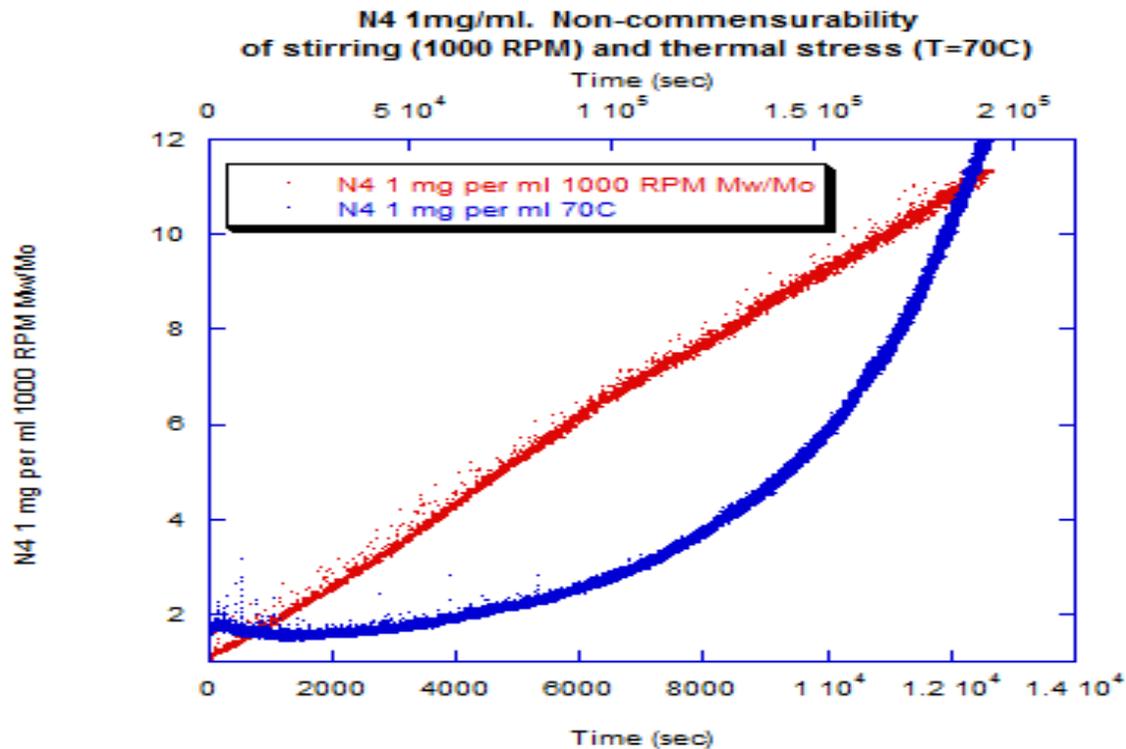
Superposable



Thermal stress: Time scaled; data superposable



# Distinguishing Kinetic Mechanisms via Superposability- Stir Stress

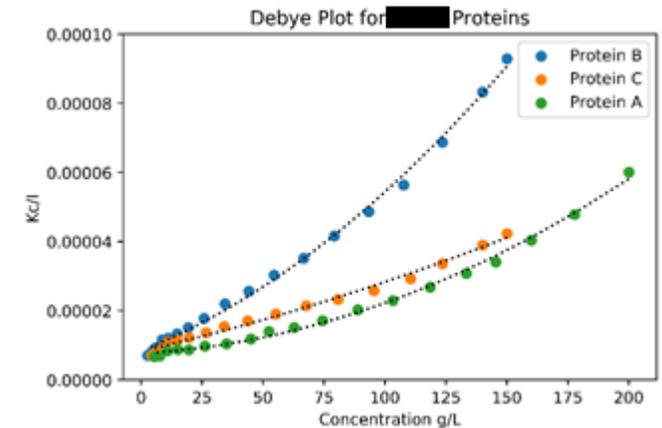
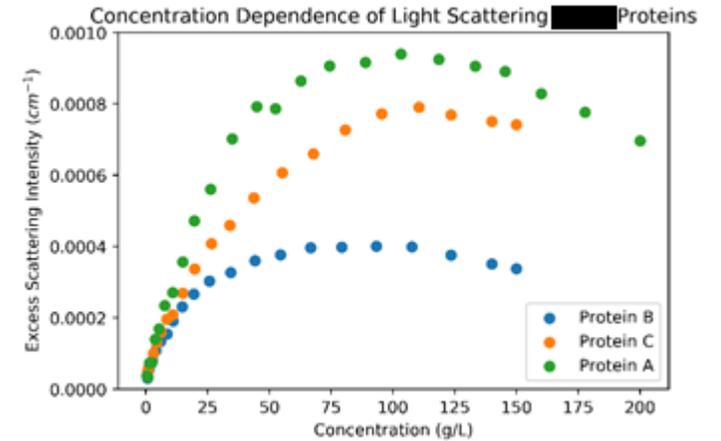


- Stirring and thermal stress with scaled timebases are not superposable
- Indicative of different kinetic mechanisms

# Generation of Debye Plots

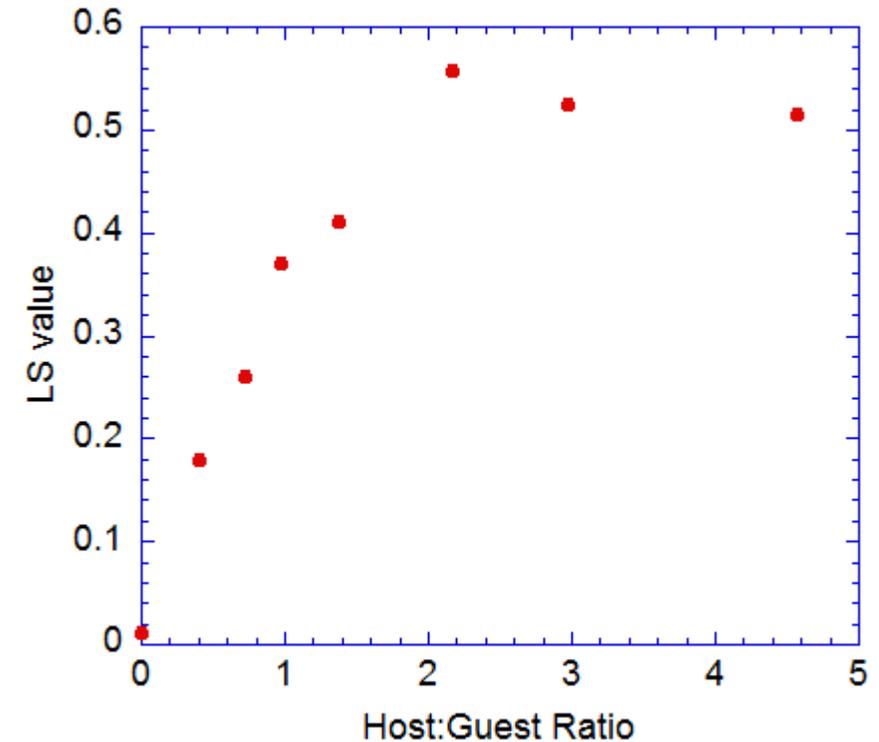
- Samples directly titrated in cell w/ gentle hand mixing
- LS v. concentration used to generate Debye plot for calculation of viral coefficients
- May require several different starting sample concentrations to cover concentration range
- Data can be collected in a single file

Debye Statistics for Proteins			
Label	$A_3$ ( $cm^2 mol g^{-2}$ )	$A_2$ ( $cm^2 mol g^{-2}$ )	$M_w$ (kDa)
Protein A	5.35e-4	1.9e-5	131
Protein B	8.95e-4	1.40e-4	119
Protein C	1.86e-4	8.2e-5	121



# Probing Supramolecular Host:Guest Interactions

- Host:Guest ratios for large molecule interactions can be determined by ARGENT
- Methodology is similar to the generation of Debye plots, i.e. direct titration of host or guest
- LS signal maxima yields host:guest stoichiometry



Payne, M; Jarand, C.; Reed, W.; Grayson, S., *Abstracts of Papers, 255th ACS National Meeting & Exposition*

# Summary

- ARGEN provides a simple and robust platform for monitoring protein aggregation by thermal and contact-stir stress
- ARGEN capabilities can be easily expanded to monitor a variety of other phenomena beyond thermal and contact-stir stress
- Examples of expanded methodology include:
  - non-contact stirring
  - peristaltic recirculation
  - monitoring filtration effects
  - examining capillary shear
  - secondary detection for monitoring aggregate formation/monomer loss
  - generation of Debye plots
  - probing host:guest interactions